J Exp Clin Med 2012;4(3):135-139



Contents lists available at SciVerse ScienceDirect

Journal of Experimental and Clinical Medicine

journal homepage: http://www.jecm-online.com

REVIEW ARTICLE

Roles of Glycogen Synthase Kinase-3 in Alzheimer's Disease: From Pathology to Treatment Target

Hsing-Cheng Liu^{1,2}, Sy-Jye Leu^{3,4,5}, De-Maw Chuang⁶*

¹ Department of General Psychiatry, Taipei City Psychiatric Center, Taipei City Hospital, Taipei, Taiwan

² Department of Psychiatry, School of Medicine, Taipei Medical University, Taipei, Taiwan

³ Department of Microbiology and Immunology, Taipei Medical University, Taipei, Taiwan

⁴ Graduate Institute of Medical Science, Taipei Medical University, Taipei, Taiwan

⁵ Center for Reproductive Medicine and Sciences, Taipei Medical University Hospital, Taipei, Taiwan

⁶ Molecular Neurobiology Section, National Institute of Mental Health, National Institutes of Health, Bethesda, Maryland, USA

ARTICLE INFO

Article history: Received: Jan 26, 2012 Revised: Mar 30, 2012 Accepted: Mar 30, 2012

KEY WORDS: Alzheimer's disease; beta-amyloid; glycogen synthase kinase-3; lithium; neurofibrillary tangle; tau Alzheimer's disease (AD) is a progressive neurodegenerative disorder with an unknown cause, and as of yet there is no effective treatment. The neuropathological hallmarks of AD include amyloid plaques and neurofibrillary tangle (NFT) deposits. There is evidentiary support for amyloid deposition being the primary influence driving AD pathogenesis, commonly referred to as the amyloid hypothesis of AD. But brain amyloid load is not correlated with AD severity; instead, NFT formation has been shown to be associated with disease progression. Therefore, advocates of the tau hypothesis strongly postulate that NFT accumulation is critical for neuronal loss and AD development. Hence, inhibition of NFT formation/accumulation is one of the treatment strategies to combat AD. NFTs consist of aggregations of paired helical hyperphosphorylated tau protein, one of the major microtubule-associated proteins. The hyperphosphorylation of tau impairs its normal maintenance function for cytoskeleton stability, and induces a toxic sequestration of normal tau and other microtubule-associated proteins. Glycogen synthase kinase-3 (GSK-3) is the main enzyme that phosphorylates tau, and an increase in GSK-3 activity has been observed in AD patients. GSK-3 inhibition by lithium, a major mood stabilizer that is used to treat bipolar disorder, has been shown to reduce tau phosphorylation and even decrease amyloid burden in the brain of AD animal models. This supports the notion of GSK-3 inhibition as a potential avenue for AD treatment.

 $Copyright @ 2012, Taipei \ Medical \ University. \ Published \ by \ Elsevier \ Taiwan \ LLC. \ All \ rights \ reserved.$

1. Introduction

Alzheimer's disease (AD) is a neurodegenerative disorder characterized by progressive and irreversible memory loss and other cognitive impairments. AD patients also suffer from various psychiatric symptoms such as psychosis, hallucinations, depression, anxiety, sleep disorder, and aberrant behavior disturbances.¹ These symptoms have a devastating emotional impact on the patients and their caregivers.² There is also a tremendous economic burden on the families and society as a whole; it is estimated that the direct cost of dementia is \$148 billion annually in the United States alone.^{3–5}

The medications used to treat the cognitive manifestations of AD include cholinesterase inhibitors (donepezil, rivastigmine, tacrine, and galantamine) and an *N*-methyl-D-aspartate (NMDA)

E-mail: D.-M. Chuang <chuang@mail.nih.gov>

receptor antagonist (memantine). The effects of these symptomatic treatment agents are limited.⁶ For psychiatric and behavioral symptoms, psychotropic agents such as second-generation (atypical) antipsychotic drugs, antidepressants, and anticonvulsants are commonly used. In addition to pharmacological treatment, non-pharmacological therapies have shown some clinical beneficial effects.⁷ However, these current therapies target only the symptoms and have no effect on delaying or halting the neurodegenerative progression of AD. Thus, the search for new treatment strategies for this incurable disease is critical.

Journal of Experimental and

Clinical Medicine

The neuropathological hallmarks of AD are the deposition of extracellular amyloid plaques and the formation of intracellular neurofibrillary tangles (NFTs). Studies of autosomal-dominant familial AD patients have implicated mutations in the following three genes in the development of AD: amyloid precursor protein (APP, on chromosome 21), presenilin 1 (PS1, on chromosome 14), and presenilin 2 (PS2, on chromosome 1).⁸ These mutations occur in genes involved in the proteolysis of APP to form β -amyloid (A β). A β is the major component of the amyloid plaque, and it is widely accepted under the amyloid hypothesis of AD that A β deposition is

^{*} Correspondence author. De-Maw Chuang, 10 Center Drive, MSC 1363, Bethesda, MD 20892-1363, USA.

^{1878-3317/\$ -} see front matter Copyright © 2012, Taipei Medical University. Published by Elsevier Taiwan LLC. All rights reserved. doi:10.1016/j.jecm.2012.04.001

the primary event in the pathogenesis of AD.^{9–11} Strategies to halt A β production and/or reduce A β deposition in the brain are believed to alter the disease progression and even cure AD.¹² Although research has provided numerous methods to reduce brain amyloid load, clinical results do not sufficiently support the amyloid hypothesis that reducing amyloid burden is beneficial for AD patients.¹³ The well-noted AN1792 A β active immunization study showed that A β burden was decreased after immunization, and yet there was no clinically significant beneficial effects observed in the immunized patients.¹⁴ Despite many explanations that have been offered to address this discrepancy, it is still reasonable to consider factors other than amyloid burden in the pathogenesis of AD.

Although the amyloid hypothesis is dominant in AD research, increasing evidence suggests that amyloid alone does not account for the AD pathology in its entirety and that NFTs, the aggregates of hyperphosphorylated tau protein, should be considered as well. Primarily, it is noted that the amyloid burden is not always correlated with clinical disease severity.^{15,16} Whereas, in contrast, NFT progression based on Braak staging resembles the clinical disease progression and NFT density is correlated with AD severity.^{17,18} These observations provide the alternative tau hypothesis of AD.¹⁹ This article examines the roles of glycogen synthase kinase-3 (GSK-3) in NFT formation and its potential as a therapeutic target for AD.

2. NFT formation in the AD brain

NFTs are aggregates of hyperphosphorylated tau in the brain. Tau is a microtubule-associated protein (MAP) expressed throughout the central nervous system (CNS), but predominantly in neuronal axons. There are six isoforms of tau protein resulting from combinations of alternative splicing of exons 2, 3, and 10. Since exon 10 constitutes one of the microtubule-binding domains, its alternative splicing would lead to isoforms containing either three or four microtubule-binding repeats (tau-3R or tau-4R, respectively).^{20–23} Partially phosphorylated tau can promote association with tubulin, which leads to microtubule stabilization. However, pathological hyperphosphorylation of tau prevents tubulin binding, thereby resulting in microtubule destabilization.²⁴ This type of microtubule disruption is believed to cause neurodegeneration. Studies have also shown that hyperphosphorylated tau can sequester normal tau and other MAPs, MAP 1 and 2.^{25,26}

The level of tau phosphorylation is regulated by a dynamic interplay between tau kinases and tau phosphatases. However, the cause of tau phosphorylation dysregulation in the AD brain is still obscure. Protein phosphatases (PPs) have broad substrate specificities: PP2A is believed to be the major tau phosphatase and is downregulated in AD brains.²⁷ About 45 phosphorylated sites have been identified in tau protein. These sites predominantly cluster in the proline-rich domain and in the C-terminal region, with few sites located within the microtubule-binding domain.^{21,28} GSK-3β was the first known tau kinase to phosphorylate tau at paired helical epitopes.²⁹ Several other kinases have also been shown to be involved in tau phosphorylation, including cyclin-dependent kinase-5 (cdk5), casein kinase-1 (ck1), and cyclic adenosine monophosphate (cAMP)-dependent protein kinase (PKA). Each kinase has its preferred serine and threonine residue(s) for tau phosphorylation. Alignment of these phosphorylated residues by each kinase with the hyperphosphorylated epitopes observed in AD brain has shown that GSK-3 β is the most important kinase to hyperphosphorylate tau.²¹ Strong evidence for GSK-3β phosphorylation of tau has been shown in an in vivo mouse brain where tau-4R wild type and P301L human tau mutant mice were crossed with GSK-3β mice.^{30,31}

In addition to phosphorylating tau, GSK-3 has the ability to affect tau splicing. GSK-3 has been shown to phosphorylate nuclear SC35, an enhancer of splicing elements that regulate exon 10 splicing in tau.^{32,33} GSK-3 is activated by A β and then phosphorylates SC35, which in turn enhances the splicing of tau exon 10, thereby decreasing the expression of tau-4R.³⁴ The imbalanced tau-3R/4R ratio enhances tau phosphorylation.³⁵ GSK-3-mediated tau splicing is interesting; however, most research investigating GSK-3 regulation of tau function has concentrated on the GSK-3-mediated phosphorylation of tau. It is believed that strategies targeting tau hyperphosphorylation could modify the NFT-induced neuro-degeneration that is associated with AD, and GSK-3 is the major target of this hypothesis.^{36–38}

Current work has suggested that aggregated tau is not the toxic factor, but rather that insoluble tau is the lethal component. The proposed toxic, insoluble tau species are thought to be oligomers, dimers, or small tau aggregates, and either remain insoluble or aggregate into larger deposits. Aberrant amyloid peptides can activate GSK-3, which in turn contributes to the hyperphosphorylation of tau. Currently, the tau hypothesis is gaining ground over the amyloid hypothesis with regard to the development of AD and disease progression.³⁹

3. GSK-3 in the CNS

GSK-3 was identified in 1980 as one of several protein kinases that phosphorylate glycogen synthase to regulate glycogen synthesis. In recent years. GSK-3 has been implicated in the phosphorylation of many protein substrates and in the regulation of several cellular events.⁴⁰ Moreover, GSK-3 acts as a central switch to receive many cellular signals including insulin, amino acids, and growth factors.⁴¹ In the 1990s, GSK-3 was found to play an important role in cell fate determination during embryogenesis. It acts in the WNT signaling pathway to phosphorylate and degrade β-catenin to inhibit gene expression.^{42,43} GSK-3 has also been demonstrated to regulate neurogenesis, neuronal polarization, and axon growth during brain development. Elevated GSK-3 activity is necessary for neuronal migration to the cortical plate, while lower GSK-3 activity is essential for maintaining progenitor neuron adherence to the subventricular zone.⁴⁴ Because of these diverse roles, it is speculated that GSK-3 is involved in many neurodevelopmental and neurodegenerative disorders.45

There are two GSK-3 isoforms, GSK-3 α and GSK-3 β , which are encoded by different genes on chromosomes 19 and 3, respectively.46,47 In contrast to other kinases, which activate their substrate, the GSK-3 isoforms inhibit their substrate activity through phosphorylation. This inhibition via phosphorylation is also seen in GSK-3 itself. Thus, phosphorylation of Ser21 in GSK-3α and of Ser9 in GSK-38 leads to decreased GSK-3 activity by inhibition of the priming phosphate site.^{48,49} But phosphorylation of the kinase domain, i.e., Tyr279 in GSK-3 α and Tyr216 in GSK-3 β , increases activity.^{50,51} These two GSK-3 isoforms share 84% sequence identity overall, but this increases to 95% in the kinase domain, indicating similar substrate specificities.⁴⁷ However, there are still several differences in these two isoforms. For example, GSK-3β deletion causes embryonic lethality, suggesting that GSK- 3α does not compensate for the loss of GSK-3 β function. 52 GSK-3 β disruption-induced embryonic lethality is likely due to severe liver degeneration, a phenotype consistent with excessive tumor necrosis factor toxicity, as observed in mice lacking activation genes of the transcription factor NF-kB. This suggests that NF-kB is regulated by GSK-3 β , but not GSK-3 α , at the level of the transcriptional complex. NF-kB activation has been suggested to be one of the differentiating factors between these two GSK-3 isoforms.⁵³ In contrast to GSK-3β deletion, GSK-3α knock-out mice survive with only aberrant behavioral abnormalities.⁵⁴ Conditional overexpression of GSK-3β in adult mouse brain increases tau phosphorylation, decreases nuclear β -catenin levels, and induces neuronal death in the hippocampus.⁵⁵ This profile resembles the tau pathology that occurs in AD. In rodents and humans, an alternative splice variant of GSK-38, GSK-382, has been reported. This variant contains a 13 amino acid insertion in an external loop near the catalytic domain. In contrast to the ubiquitously expressed GSK-3\beta1, GSK-3\beta2 is expressed specifically in the nervous system, and the highest levels are found during development.⁵⁶ GSK-3 β 2 has lower tau phosphorylation activity than GSK-3 β 1.⁵⁷ In regard to neuron-specific functions, GSK-3^β has been reported to mediate two synaptic activities: NMDA receptor-mediated long-term potentiation (LTP) and long-term depression (LTD). GSK-3β Ser9 phosphorylation by the PI3K/AKT signaling, which inhibits GSK-3 β , is crucial for the formation of LTP or LTD that regulates synaptic plasticity and cognition.^{58–60} Overall, GSK-3 β has been studied more extensively than GSK-3a, and its proposed role in AD pathophysiology has been of greater interest.

4. Dysregulation of GSK-3 in the AD brain

Direct evidence from human AD brains provides insight into the role of GSK-3 in the pathophysiology of this disorder. It has been shown that the total amounts of both GSK-3 α and GSK-3 β are decreased in human AD brain extracts irrespective of their activities.⁶¹ Studies have demonstrated that there is a significant increase in the level of pTvr216 GSK-38 (active form) in the frontal cortex of AD patients. Moreover, there is broad somatodendritic accumulation of pTyr216 GSK-3β at all stages of neurodegeneration, but no accumulation of the inactive form of pSer9 GSK-3⁶² This suggests that activation of GSK-3 β is an early event that precedes and accompanies the formation of NFTs. Another study showed that pSer9 GSK-3β (inactive form) is co-localized with NFTs, dystrophic neurites of senile plaques, and neuropil threads. This sequestration of the phosphorylated form of GSK-3 β may be a compensatory process to prevent further tauopathy.⁶³ In a transgenic AD animal study, GSK-3 activity was increased prior to amyloid deposition, suggesting that amyloid induces tauopathy through GSK-3 activation.³¹ These human and animal studies support the notion that increased GSK-3β activity may play an important role in AD pathogenesis, although further work is still needed to solidify this notion. It should also be noted that regulation of GSK- $3\alpha/\beta$ activity through phosphorylation of Ser21/Ser9 (inhibition) and/or Tyr279/ Tyr216 (activation) is still a matter of controversy, not only in AD, but in normal physiology as well.

5. Effects of GSK-3 inhibition in experimental AD models

Several GSK-3 inhibition studies in AD animal models have shown promising results for both pathology and behavior. In an intracerebroventricular A β infusion AD model, co-infusion of SB216763, an ATP-competitive GSK-3 inhibitor, normalizes A β -induced elevations of phospho-tau, caspase-3, phospho-c-jun *N*-terminal kinase, neuronal DNA fragmentation, and gliosis.⁶⁴ Lithium, an anti-bipolar drug and well-known GSK-3 inhibitor,⁶⁵ has been shown to decrease tau phosphorylation through GSK-3 inhibition in AD animals,^{66,67} and even to downregulate tau transcription in cultured cortical neurons.⁶⁸ Lithium also increases PP2A activity,⁶⁹ while decreasing tau phosphorylation.⁶⁸

In addition to modulating tau phosphorylation, GSK-3 is also involved in the APP/A β processing. In a *Drosophila* AD model, lithium attenuates A β 42-induced fly locomotor activity changes and prolongs life span of the A β 42 overexpressing flies.⁷⁰ Axonal transport deficits are also reversed in this model.⁷¹ In tauopathy mouse models, lithium inhibits GSK-3-mediated phosphorylation of tau and the associated neuronal degeneration,⁷² as well as promotes ubiquitination, which results in a decrease of tau lesions.⁷³ Furthermore, lithium was found to prevent the development of NFTs in a mouse model developed to display advanced neurofibrillary pathology,74 and to decrease mutant tau protein aggregation in another mouse transgenic model.⁶⁶ An additional interesting finding is that AB42 levels are reduced after GSK-3 inhibition, suggesting a direct role of GSK-3 in the regulation of Aβ42 biogenesis.⁷⁰ In transgenic mouse models of AD, lithium also improves cognition and reduces A β burden.^{75,76} Additionally, previous studies have demonstrated that GSK-3^β phosphorylates the intracellular domain of APP protein,⁷⁷ and GSK-3 α interacts with presenilin of the γ -secretase complex to interfere in APP processing.⁷⁸ Lithium has also shown promising results in another AD transgenic mouse model overexpressing APP where the treatment decreases tau hyperphosphorylation and Aβ burden, reduces neurodegeneration in the cortex and hippocampus, and even normalizes memory deficits in a water maze test.⁷⁹ Figure 1 is a schematic illustration of the multiple roles of GSK-3 in mediating tau hyperphosphorylation and $A\beta$ generation, and shows that GSK-3 can serve as a potential target of AD treatment.

There are few studies looking at GSK-3 inhibition in humans. A registry database study in Denmark showed that continuous lithium treatment is beneficial in lowering dementia risk.⁸⁰ In a 10-week trial for AD patients, the lithium treatment group did not show cognitive improvements or changes in cerebrospinal fluid (CSF) biomarkers.⁸¹ Another lithium trial was tested in mild cognitively impaired patients for a treatment period that was



Figure 1 Proposed roles of GSK-3 in the pathophysiology of AD. GSK-3 is proposed to have multiple actions in the genesis of AD pathology. It has been shown that GSK-3 is involved in the mRNA splicing of Tau protein, producing the tau-3R/4R isoforms, GSK-3 is a major kinase that phosphorylates Tau protein at multiple sites. Hyperactive GSK-3 has been detected in the brain of AD patients, thus resulting in tau hyperphosphorylation, which triggers neurofibrillary tangle formation and microtubule instability. These effects play a major role in the neuronal death associated with AD pathology. On the other hand, APP is processed by the enzymatic reactions of β - and γ secretases, producing $A\beta$ and ultimately leading to the formation of an amyloid plaque. The APP-AB pathway also contributes to the neuronal death in the affected brain areas. A β itself can further activate GSK-3, inducing a vicious cycle of tau mRNA splicing and protein phosphorylation. GSK-3 has been shown to facilitate APP processing by regulating the activity of the γ -secretase complex. Lithium and other GSK-3 inhibitors have been shown to reduce tau hyperphosphorylation and Aß generation as well as to improve learning and memory in transgenic mouse models of AD. They have also been used in clinical trials to treat AD patients. Arrows represent stimulatory connections, and lines with flattened ends represent inhibitory connections. $A\beta=\beta$ amyloid; AD = Alzheimer's disease; APP = amyloid precursor protein; GSK-3 = glycogen synthase kinase-3; tau-3R/4R = tau isoforms containing either three or four microtubule-binding repeats.

extended to 12 months. Lithium treatment is associated with a significant decrease in CSF concentrations of phosphorylated tau and improved cognitive performance.⁸² This human study suggests that GSK-3 inhibition by lithium may be beneficial if the treatment begins at an earlier stage and is maintained for a longer period. Lithium and other GSK-3 inhibitors are also under Phase II clinical trials for tauopathies. Lithium is undergoing testing at the University of Sao Paulo for AD; Tideglusib (NP-12) is under testing by Noscira for AD and progressive supranuclear palsy (PSP). Valproic acid, a histone deacetylase inhibitor with GSK-3 inhibitory activity, is undergoing Phase II trials at Nantes University Hospital for PSP.⁸³

The current trend of AD treatment is to focus on the neuropathological hallmarks of AD: A β and tau. GSK-3 plays a major role in tau hyperphosphorylation and also mediates APP processing in part. Although there are few studies investigating GSK-3 inhibition for AD in humans, we still believe in its therapeutic potential, especially if more effective and safer GSK-3 inhibitors are discovered, and a plethora of chemical compounds have already been tested for their GSK-3 inhibitory activities.⁸⁴ We concede that AD treatment could require other avenues than GSK-3 inhibition; however, we still expect GSK-3 inhibition to be an important aspect for AD treatment.

Acknowledgments

The authors wish to thank the Department of Health, Taipei City Government (96001-62-031); the Department of Health, Taipei City Government (96001-62-037); the Department of Health, Taipei City Government (99001-62-039); and the Intramural Research Program of NIMH, NIH, USA, for their support. The excellent editorial assistance from Peter Leeds and Fairouz Chibane of NIMH, NIH, is greatly appreciated.

References

- Ballard CG, Gauthier S, Cummings JL, Brodaty H, Grossberg GT, Robert P, Lyketsos CG. Management of agitation and aggression associated with Alzheimer disease. Nat Rev Neurol 2009;5:245–55.
- Mohamed S, Rosenheck R, Lyketsos CG, Schneider LS. Caregiver burden in Alzheimer disease: cross-sectional and longitudinal patient correlates. Am J Geriatr Psychiatry 2010;18:917–27.
- Rojas G, Bartoloni L, Dillon C, Serrano CM, Iturry M, Allegri RF. Clinical and economic characteristics associated with direct costs of Alzheimer's, frontotemporal and vascular dementia in Argentina. *Int Psychogeriatr* 2011;23: 554–61.
- Cappell J, Herrmann N, Cornish S, Lanctot KL. The pharmacoeconomics of cognitive enhancers in moderate to severe Alzheimer's disease. CNS Drugs 2010;24:909–27.
- Alzheimer's Association. 2009 Alzheimer's disease facts and figures. Alzheimers Dement 2009;5:234–70.
- Hansen RA, Gartlehner G, Webb AP, Morgan LC, Moore CG, Jonas DE. Efficacy and safety of donepezil, galantamine, and rivastigmine for the treatment of Alzheimer's disease: a systematic review and meta-analysis. *Clin Interv Aging* 2008;3:211–25.
- Olazaran J, Reisberg B, Clare L, Cruz I, Pena-Casanova J, Del Ser T, Woods B, et al. Nonpharmacological therapies in Alzheimer's disease: a systematic review of efficacy. *Dement Geriatr Cogn Disord* 2010;**30**:161–78.
- Bekris LM, Yu CE, Bird TD, Tsuang DW. Genetics of Alzheimer disease. J Geriatr Psychiatry Neurol 2010;23:213–27.
- 9. Hardy J, Allsop D. Amyloid deposition as the central event in the aetiology of Alzheimer's disease. *Trends Pharmacol Sci* 1991;**12**:383–8.
- Hardy J, Selkoe DJ. The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science* 2002;297:353–6.
- 11. Hardy JA, Higgins GA. Alzheimer's disease: the amyloid cascade hypothesis. *Science* 1992;**256**:184–5.
- Carter MD, Simms GA, Weaver DF. The development of new therapeutics for Alzheimer's disease. Clin Pharmacol Ther 2010;88:475–86.
- Hardy J. The amyloid hypothesis for Alzheimer's disease: a critical reappraisal. J Neurochem 2009;110:1129–34.
- Holmes C, Boche D, Wilkinson D, Yadegarfar G, Hopkins V, Bayer A, Jones RW, et al. Long-term effects of Abeta42 immunisation in Alzheimer's disease: follow-up of a randomised, placebo-controlled phase I trial. *Lancet* 2008;**372**: 216–23.

- Josephs KA, Whitwell JL, Ahmed Z, Shiung MM, Weigand SD, Knopman DS, Boeve BF, et al. Beta-amyloid burden is not associated with rates of brain atrophy. *Ann Neurol* 2008;63:204–12.
- Villemagne VL, Pike KE, Chetelat G, Ellis KA, Mulligan RS, Bourgeat P, Ackermann U, et al. Longitudinal assessment of Abeta and cognition in aging and Alzheimer disease. Ann Neurol 2011;69:181–92.
- Braak H, Braak E. Staging of Alzheimer's disease-related neurofibrillary changes. *Neurobiol Aging* 1995;16:271–8. discussion 8–84.
- Giannakopoulos P, Herrmann FR, Bussiere T, Bouras C, Kovari E, Perl DP, Morrison JH, et al. Tangle and neuron numbers, but not amyloid load, predict cognitive status in Alzheimer's disease. *Neurology* 2003;60:1495–500.
- Maccioni RB, Farias G, Morales I, Navarrete L. The revitalized tau hypothesis on Alzheimer's disease. Arch Med Res 2010;41:226–31.
- Ballard C, Khan Z, Clack H, Corbett A. Nonpharmacological treatment of Alzheimer disease. Can J Psychiatry 2011;56:589–95.
- Hanger DP, Anderton BH, Noble W. Tau phosphorylation: the therapeutic challenge for neurodegenerative disease. *Trends Mol Med* 2009;15:112–9.
- Liu F, Gong CX. Tau exon 10 alternative splicing and tauopathies. Mol Neurodegener 2008;3:8.
- Zhou J, Yu Q, Zou T. Alternative splicing of exon 10 in the tau gene as a target for treatment of tauopathies. *BMC Neurosci* 2008;9(Suppl. 2):S10.
- Drechsel DN, Hyman AA, Cobb MH, Kirschner MW. Modulation of the dynamic instability of tubulin assembly by the microtubule-associated protein tau. *Mol Biol Cell* 1992;3:1141-54.
- Alonso AD, Zaidi T, Novak M, Barra HS, Grundke-Iqbal I, Iqbal K. Interaction of tau isoforms with Alzheimer's disease abnormally hyperphosphorylated tau and in vitro phosphorylation into the disease-like protein. J Biol Chem 2001;276:37967–73.
- 26. Alonso AD, Grundke-Iqbal I, Barra HS, Iqbal K. Abnormal phosphorylation of tau and the mechanism of Alzheimer neurofibrillary degeneration: sequestration of microtubule-associated proteins 1 and 2 and the disassembly of microtubules by the abnormal tau. *Proc Natl Acad Sci USA* 1997;**94**:298–303.
- Liu F, Grundke-Iqbal I, Iqbal K, Gong CX. Contributions of protein phosphatases PP1, PP2A, PP2B and PP5 to the regulation of tau phosphorylation. *Eur J Neurosci* 2005;**22**:1942–50.
- Morishima-Kawashima M, Hasegawa M, Takio K, Suzuki M, Yoshida H, Watanabe A, Titani K, et al. Hyperphosphorylation of tau in PHF. *Neurobiol Aging* 1995;16:365-71. discussion 71-80.
- Ishiguro K, Shiratsuchi A, Sato S, Omori A, Arioka M, Kobayashi S, Uchida T, et al. Glycogen synthase kinase 3 beta is identical to tau protein kinase I generating several epitopes of paired helical filaments. *FEBS Lett* 1993;**325**:167–72.
- Spittaels K, Van den Haute C, Van Dorpe J, Geerts H, Mercken M, Bruynseels K, Lasrado R, et al. Glycogen synthase kinase-3beta phosphorylates protein tau and rescues the axonopathy in the central nervous system of human fourrepeat tau transgenic mice. J Biol Chem 2000;275:41340-9.
- Terwel D, Muyllaert D, Dewachter I, Borghgraef P, Croes S, Devijver H, Van Leuven F. Amyloid activates GSK-3beta to aggravate neuronal tauopathy in bigenic mice. Am J Pathol 2008; 172:786–98.
- D'Souza I, Schellenberg GD. Determinants of 4-repeat tau expression. Coordination between enhancing and inhibitory splicing sequences for exon 10 inclusion. J Biol Chem 2000;275:17700–9.
- Hernandez F, Perez M, Lucas JJ, Mata AM, Bhat R, Avila J. Glycogen synthase kinase-3 plays a crucial role in tau exon 10 splicing and intranuclear distribution of SC35. Implications for Alzheimer's disease. J Biol Chem 2004;279:3801–6.
- Chen KL, Yuan RY, Hu CJ, Hsu CY. Amyloid-beta peptide alteration of tau exon-10 splicing via the GSK3beta-SC35 pathway. *Neurobiol Dis* 2010;40:378-85.
- Schweers O, Mandelkow EM, Biernat J, Mandelkow E. Oxidation of cysteine-322 in the repeat domain of microtubule-associated protein tau controls the in vitro assembly of paired helical filaments. *Proc Natl Acad Sci USA* 1995;92:8463–7.
- Gong CX, Iqbal K. Hyperphosphorylation of microtubule-associated protein tau: a promising therapeutic target for Alzheimer disease. *Curr Med Chem* 2008;15:2321–8.
- Iqbal K, Grundke-Iqbal I. Alzheimer neurofibrillary degeneration: significance, etiopathogenesis, therapeutics and prevention. J Cell Mol Med 2008;12:38–55.
- Martinez A, Perez DI. GSK-3 inhibitors: a ray of hope for the treatment of Alzheimer's disease? J Alzheimers Dis 2008;15:181–91.
- 39. Kremer A, Louis JV, Jaworski T, Van Leuven F. GSK3 and Alzheimer's disease: facts and fiction. Front Mol Neurosci 2011;4:17.
- Frame S, Cohen P. GSK3 takes centre stage more than 20 years after its discovery. *Biochem* J 2001;359:1–16.
- 41. Cohen P, Frame S. The renaissance of GSK3. Nat Rev Mol Cell Biol 2001;2: 769-76.
- Zeng L, Fagotto F, Zhang T, Hsu W, Vasicek TJ, Perry III WL, Lee JJ, et al. The mouse fused locus encodes axin, an inhibitor of the Wnt signaling pathway that regulates embryonic axis formation. *Cell* 1997;**90**:181–92.
- Ikeda S, Kishida S, Yamamoto H, Murai H, Koyama S, Kikuchi A. Axin, a negative regulator of the Wnt signaling pathway, forms a complex with GSK-3beta and beta-catenin and promotes GSK-3beta-dependent phosphorylation of betacatenin. *EMBO J* 1998;17:1371–84.
- Hur EM, Zhou FQ. GSK3 signalling in neural development. Nat Rev Neurosci 2010;11:539–51.
- Chiu C-T, Chuang D-M. Molecular actions and therapeutic potential of lithium in preclinical and clinical studies of CNS disorders. *Pharmacol Therap* 2010;**128**: 281–304.

- 46. Shaw PC, Davies AF, Lau KF, Garcia-Barcelo M, Waye MM, Lovestone S, Miller CC, et al. Isolation and chromosomal mapping of human glycogen synthase kinase-3 alpha and -3 beta encoding genes. *Genome* 1998;41:720–7.
- Woodgett JR. Molecular cloning and expression of glycogen synthase kinase-3/ factor A. EMBO J 1990;9:2431–8.
- Frame S, Cohen P, Biondi RM. A common phosphate binding site explains the unique substrate specificity of GSK3 and its inactivation by phosphorylation. *Mol Cell* 2001;7:1321–7.
- ter Haar E, Coll JT, Austen DA, Hsiao HM, Swenson L, Jain J. Structure of GSK3beta reveals a primed phosphorylation mechanism. *Nat Struct Biol* 2001;8:593–6.
- Hughes K, Nikolakaki E, Plyte SE, Totty NF, Woodgett JR. Modulation of the glycogen synthase kinase-3 family by tyrosine phosphorylation. *EMBO J* 1993;12:803-8.
- Wang QM, Fiol CJ, DePaoli-Roach AA, Roach PJ. Glycogen synthase kinase-3 beta is a dual specificity kinase differentially regulated by tyrosine and serine/threonine phosphorylation. J Biol Chem 1994;269:14566-74.
- Hoeflich KP, Luo J, Rubie EA, Tsao MS, Jin O, Woodgett JR. Requirement for glycogen synthase kinase-3beta in cell survival and NF-kappaB activation. *Nature* 2000:406:86–90.
- Ali A, Hoeflich KP, Woodgett JR. Glycogen synthase kinase-3: properties, functions, and regulation. *Chem Rev* 2001;**101**:2527–40.
- Kaidanovich-Beilin O, Lipina TV, Takao K, van Eede M, Hattori S, Laliberte C, Khan M, et al. Abnormalities in brain structure and behavior in GSK-3alpha mutant mice. *Mol Brain* 2009;2:35.
- Lucas JJ, Hernandez F, Gomez-Ramos P, Moran MA, Hen R, Avila J. Decreased nuclear beta-catenin, tau hyperphosphorylation and neurodegeneration in GSK-3beta conditional transgenic mice. *EMBO J* 2001;20:27–39.
- Mukai F, Ishiguro K, Sano Y, Fujita SC. Alternative splicing isoform of tau protein kinase l/glycogen synthase kinase 3beta. J Neurochem 2002;81: 1073–83.
- Saeki K, Machida M, Kinoshita Y, Takasawa R, Tanuma S. Glycogen synthase kinase-3beta2 has lower phosphorylation activity to tau than glycogen synthase kinase-3beta1. *Biol Pharm Bull* 2011;34:146–9.
- Hooper C, Markevich V, Plattner F, Killick R, Schofield E, Engel T, Hernandez F, et al. Glycogen synthase kinase-3 inhibition is integral to long-term potentiation. *Eur J Neurosci* 2007;25:81–6.
- Peineau S, Bradley C, Taghibiglou C, Doherty A, Bortolotto ZA, Wang YT, Collingridge GL. The role of GSK-3 in synaptic plasticity. Br J Pharmacol 2008;153(Suppl. 1):S428–37.
- Peineau S, Taghibiglou C, Bradley C, Wong TP, Liu L, Lu J, Lo E, et al. LTP inhibits LTD in the hippocampus via regulation of GSK3beta. *Neuron* 2007;53:703–17.
- Baum L, Hansen L, Masliah E, Saitoh T. Glycogen synthase kinase 3 alteration in Alzheimer disease is related to neurofibrillary tangle formation. *Mol Chem Neuropathol* 1996;29:253–61.
- Leroy K, Yilmaz Z, Brion JP. Increased level of active GSK-3beta in Alzheimer's disease and accumulation in argyrophilic grains and in neurones at different stages of neurofibrillary degeneration. *Neuropathol Appl Neurobiol* 2007;**33**: 43–55.
- 63. Ferrer I, Barrachina M, Puig B. Glycogen synthase kinase-3 is associated with neuronal and glial hyperphosphorylated tau deposits in Alzheimer's disease, Pick's disease, progressive supranuclear palsy and corticobasal degeneration. *Acta Neuropathol* 2002;**104**:583–91.
- 64. Hu S, Begum AN, Jones MR, Oh MS, Beech WK, Beech BH, Yang F, et al. GSK3 inhibitors show benefits in an Alzheimer's disease (AD) model of neurodegeneration but adverse effects in control animals. *Neurobiol Dis* 2009;**33**: 193–206.
- Stambolic V, Ruel L, Woodgett JR. Lithium inhibits glycogen synthase kinase-3 activity and mimics wingless signalling in intact cells. Curr Biol 1996;6:1664–8.

- Perez M, Hernandez F, Lim F, Diaz-Nido J, Avila J. Chronic lithium treatment decreases mutant tau protein aggregation in a transgenic mouse model. J Alzheimers Dis 2003;5:301–8.
- 67. Caccamo A, Oddo S, Tran LX, LaFerla FM. Lithium reduces tau phosphorylation but not Abeta or working memory deficits in a transgenic model with both plaques and tangles. *Am J Pathol* 2007;**170**:1669–75.
- Rametti A, Esclaire F, Yardin C, Terro F. Linking alterations in tau phosphorylation and cleavage during neuronal apoptosis. J Biol Chem 2004;279:54518–28.
- Tsuji S, Morinobu S, Tanaka K, Kawano K, Yamawaki S. Lithium, but not valproate, induces the serine/threonine phosphatase activity of protein phosphatase 2A in the rat brain, without affecting its expression. J Neural Transm 2003;110:413–25.
- Sofola O, Kerr F, Rogers I, Killick R, Augustin H, Gandy C, Allen MJ, et al. Inhibition of GSK-3 ameliorates Abeta pathology in an adult-onset Drosophila model of Alzheimer's disease. PLoS Genet 2010;6(9):e1001087.
- Mudher A, Shepherd D, Newman TA, Mildren P, Jukes JP, Squire A, Mears A, et al. GSK-3beta inhibition reverses axonal transport defects and behavioural phenotypes in Drosophila. *Mol Psychiatry* 2004;9:522–30.
- Noble W, Planel E, Zehr C, Olm V, Meyerson J, Suleman F, Gaynor K, et al. Inhibition of glycogen synthase kinase-3 by lithium correlates with reduced tauopathy and degeneration in vivo. Proc Natl Acad Sci USA 2005;102:6990–5.
- Nakashima H, Ishihara T, Suguimoto P, Yokota O, Oshima E, Kugo A, Terada S, et al. Chronic lithium treatment decreases tau lesions by promoting ubiquitination in a mouse model of tauopathies. *Acta Neuropathol* 2005;**110**:547–56.
- Leroy K, Ando K, Heraud C, Yilmaz Z, Authelet M, Boeynaems JM, Buee L, et al. Lithium treatment arrests the development of neurofibrillary tangles in mutant tau transgenic mice with advanced neurofibrillary pathology. J Alzheimers Dis 2010;19:705–19.
- Toledo EM, Inestrosa NC. Activation of Wnt signaling by lithium and rosiglitazone reduced spatial memory impairment and neurodegeneration in brains of an APPswe/PSEN1DeltaE9 mouse model of Alzheimer's disease. *Mol Psychiatry* 2010;15:272–85.
- Zhang X, Heng X, Li T, Li L, Yang D, Du Y, Doody RS, et al. Long-term treatment with lithium alleviates memory deficits and reduces amyloid-beta production in an aged Alzheimer's disease transgenic mouse model. J Alzheimers Dis 2011;24:739–49.
- Aplin AE, Gibb GM, Jacobsen JS, Gallo JM, Anderton BH. *In vitro* phosphorylation of the cytoplasmic domain of the amyloid precursor protein by glycogen synthase kinase-3beta. *J Neurochem* 1996;67:699–707.
- Phiel CJ, Wilson CA, Lee VM, Klein PS. GSK-3alpha regulates production of Alzheimer's disease amyloid-beta peptides. *Nature* 2003;423:435–9.
- Rockenstein E, Torrance M, Adame A, Mante M, Bar-on P, Rose JB, Crews L, et al. Neuroprotective effects of regulators of the glycogen synthase kinase-3beta signaling pathway in a transgenic model of Alzheimer's disease are associated with reduced amyloid precursor protein phosphorylation. J Neurosci 2007;27:1981–91.
- Kessing LV, Sondergard L, Forman JL, Andersen PK. Lithium treatment and risk of dementia. Arch Gen Psychiatry 2008;65:1331–5.
- Hampel H, Ewers M, Burger K, Annas P, Mortberg A, Bogstedt A, Frolich L, et al. Lithium trial in Alzheimer's disease: a randomized, single-blind, placebocontrolled, multicenter 10-week study. J Clin Psychiatry 2009;70:922–31.
- Forlenza OV, Diniz BS, Radanovic M, Santos FS, Talib LL, Gattaz WF. Disease-modifying properties of long-term lithium treatment for amnestic mild cognitive impairment: randomised controlled trial. Br J Psychiatry 2011;198: 351–6.
- Medina M, Garrido JJ, Wandosell FG. Modulation of GSK-3 as a therapeutic strategy on tau pathologies. Front Mol Neurosci 2011;4:24.
- Cohen P, Goedert M. GSK3 inhibitors: development and therapeutic potential. Nat Rev Drug Discov 2004;3:479-87.